

APPENDIX A

Sample Benchsheets

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Facility Name: _____

BOD₅ Benchsheet

Sample Location (specific) _____

Sample Type (grab, xx hr Comp., etc.) _____

Raw _____
 Final _____

Sample Date: _____

Test Date: _____

Collected by: _____

Analyst: _____

Samples IN Date: _____

Samples OUT Date: _____

Time: _____

Time: _____

Room Temp (°C) _____

Room Temp (°C) _____

Barometric pressure _____

Barometric pressure _____

	Bottle #	Sample mLs	Initial DO (mg/L)	Final DO (mg/L)	Depletion (mg/L)	Comments
Blank						Depletion must be less than 0.2 mg/L
Seeded						
Blank						

Seed source: _____

Seed Correction factor (SCF): _____

Sample	Bottle #	Sample mLs	Seed mLs added	Initial DO	Final DO	DO depletion	SCF	Dilution factor	BOD ₅ mg/L	¹ Average BOD ₅
		A		B	C	D=B-C	E	F=300/A	F x (D-E)	
Raw										
Final										
GGA										
Replicate of _____										

¹ Average only those values which are associated with a depletion of at least 2 mg/L and a final DO ≥ 1 mg/L.

Calculation = BOD₅ mg/L = [(B-C)-E] x F

Facility Name: _____

TSS Benchsheet

Sample Location (specific) _____

Raw _____

Final _____

Sample Type (grab, xx hr Comp., etc.) _____

Sample Date: _____

Collected by: _____

Test Date: _____

Analyst: _____

Samples IN Date: _____

Time: _____

Samples OUT Date: _____

Time: _____

Oven Temp (°C) _____

Oven Temp (°C) _____

		Influent (Raw)	Effluent (Final)	Replicate (of _____)	Other: _____	Other: _____
Crucible/filter ID						
¹ Volume Filtered (mLs)						
Crucible/Filter AFTER drying (g) or (mg)	1 st weight					
	2 nd weight					
	3 rd weight					
Crucible/Filter tare weight (g) or (mg)	1 st weight					
	2 nd weight					
	3 rd weight					
³ Weight of dry solids (mg)						
⁴ TSS (mg/L)						

¹ Filter sufficient volume of sample to capture at least 1 mg (0.001 g) of solids.

² If samples are dried overnight, then re-drying/re-weighing is not necessary. Otherwise, at least once each quarter, one sample must be dried/weighed, and the re-dried and re-weighed to demonstrate that a constant dry weight is achieved based on the drying time employed.

³ Milligrams (mg) = grams x 100. The weight of dry solids must be less than 200 milligrams (0.2 g) or the analysis must be repeated using an appropriately smaller volume.

⁴ TSS = $\frac{\text{weight of dry solids (mg)}}{\text{volume Filtered (mL)}} \times 1,000$

Facility Name: _____

Ammonia (by electrode) Benchsheet

Sample Location (specific) _____

Raw _____

Final _____

Sample Type (grab, xx hr Comp., etc.) _____

Sample Date: _____

Test Date: _____

Collected by: _____

Analyst: _____

Calibration by: Internal (Direct Read): _____

Semi-logarithmic paper: _____

Relative millivolts: _____

Linear regression: _____

For **all** calibrations- Slope (per decade of concentration) = _____ (*must be within -54 to -60 mV*)

For linear regressions- Intercept = _____

correlation coefficient (r) = _____ (*must be ≥ 0.995*)

Concentration = Inverse/antilog of [sample mV x *slope* + *intercept*]

For ALL calibrations

~~For linear regressions~~

Standard Concentration (mg/L)	Millivolts (mV)	Log ₁₀ of concentration ¹ (mg/L)	Regression concentration ² (mg/L)
Blank			

ALL calibrations must be based on at least 3 standards

¹ Take the log of the concentration

² obtained by solving for concentration using the millivolts of the standards

	Influent (Raw)	Effluent (Final)	Replicate (of _____)	Matrix Spike (of _____)
Distilled? (Y/N)				
Sample Volume mLs				
Dilution Factor (DF)				
Millivolts (mV)				
* mg/L from calibration				
** Final mg/L as NH ₃				

* This is the concentration determined directly from the calibration. If there was no dilution involved (i.e., you used the same volume of sample as was used for the standards, then this is also equal to the final ammonia concentration.

** Final concentration = mg/L from calibration X DF

Dilution Factor = $\frac{\text{mLs used for standards}}{\text{mLs used for sample}}$

Facility Name: _____

Total Phosphorus Benchsheet

Sample Location (specific) _____

Sample Type (grab, xx hr Comp., etc.) _____

Raw _____

Final _____

Sample Date: _____

Test Date: _____

Collected by: _____

Analyst: _____

Color Development in Method Blank? (Y/N): _____

Calibration by: Internal (Direct Read): _____

Graph paper: _____

Linear regression: _____

For **all** calibrations- Slope (per decade of concentration) = _____ (*monitor for consistency or significant changes*)

Intercept = _____ (*must be less than the LOD*)

For linear regressions- correlation coefficient (r) = _____ (*must be ≥ 0.995*)

Concentration mg/L = [sample absorbance X *slope* + *intercept*]

For ALL calibrations

~~For linear regressions~~

Standard Concentration (mg? or mg/L?)	Absorbance @ 880 nm	Regression concentration ¹ (mg? or mg/L)
Blank		

ALL calibrations must be based on at least 3 standards

¹ obtained by solving for concentration using the absorbance of the standards

	Known Standard If no full calibration this day	Influent (Raw)	Effluent (Final)	Replicate (of _____)	Matrix Spike (of _____)
Sample Volume mLs					
Absorbance (<i>after coloring</i>)					
Absorbance (<i>before coloring</i>)					
Net Absorbance					
Dilution Factor (DF)					
* mg/L from calibration					
** Final mg/L as P					

* This is the concentration determined directly from the calibration. If there was no dilution involved (i.e., you used the same volume of sample as was used for the standards, then this is also equal to the final phosphorus concentration.

** Final concentration = $\frac{\text{mg from calibration}}{\text{L}} \times \text{DF}$

Dilution Factor = $\frac{\text{mLs digested for standards}}{\text{mLs digested for sample}} \times \frac{\text{mLs colored for standards}}{\text{mLs colored for sample}}$

Facility Name: _____

Residual Chlorine Benchsheet

Sample Location (specific) _____

Sample Type: GRAB

Sample Date/Time: _____

Collected by: _____

Test Date: _____

Analyst: _____

Calibration by: Internal (Direct Read): _____

Semi-logarithmic paper: _____

Relative millivolts: _____

Linear regression: _____

For **all** calibrations- Slope (per decade of concentration) = _____ (*must be within -54 to -60 mV*)

For linear regressions- Intercept = _____

correlation coefficient (r) = _____ (*must be ≥ 0.995*)

Concentration = Inverse/antilog of [sample mV x *slope + intercept*]

For ALL calibrations

For linear regressions

Standard Cl ₂ Equivalent (mg/L)	Absorbance @ 515 nm colorimetric test	Millivolts (mV) electrode method	Log ₁₀ of concentration ¹ (mg/L)	Regression concentration ² (mg/L)
Blank				

ALL calibrations should be based on at least 3 standards

¹ Take the log of the concentration

² obtained by solving for concentration using the millivolts/absorbance of the standards

	Effluent (Final)	Other: _____
Absorbance		
Millivolts (mV)		
Dilution Factor (DF)		
* mg/L from calibration		
** Final mg/L as Cl ₂		

* This is the concentration determined directly from the calibration. If there was no dilution involved (i.e., you used the same volume of sample as was used for the standards, then this is also equal to the final ammonia concentration.

** Final concentration = mg/L from calibration X DF

Dilution Factor =
$$\frac{\text{mLs used for standards}}{\text{mLs used for sample}}$$

Facility Name: _____

Fecal Coliform Benchsheet

Sample Location (specific) _____

Sample Type: GRAB

Sample Date/Time: _____

Collected by: _____

Test Date: _____

Analyst: _____

Samples IN Date: _____

Time: _____

Oven Temp (°C) _____

Samples OUT Date: _____

Time: _____

Oven Temp (°C) _____

Plate #	Sample Size (mLs)	# colonies on plate	Fecal coliforms per 100 mLs	Avg. Fecal coliforms per 100 mLs

Calculation: Fecal coliforms per 100 mL=

$$\frac{\text{\# colonies per plate}}{\text{Sample size (mL)}} \times 100$$

Lab Equipment Maintenance and Calibration Log

Month _____ Year _____

Date	Analyst Initials	Sampler Temp. (°C)	Refrig. Temp. (°C)	TSS Oven Temp °C	BOD Incubator Temp °C	Fecal Incubator Temp °C	pH Meter buffers	BOD Barometer reading	BOD Room Temp °C		
1											
2											
3											
4											
5											
6											
7											
8											
9											
10											
11											
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31											

Monthly Balance Check weight in milligram range: Actual weight(mg): _____ Measured weight(mg) _____

Monthly Balance Check weight in gram range: Actual weight (g): _____ Measured weight (g) _____

Membrane changes:

DO probe

Ammonia probe

Other probe

TSS Oven should be 103-105 °C

BOD Incubator should be 20 ± 1 C

Sampler and Refrigerator Temperature should not exceed 4 ° C (and samples should not be frozen)

pH calibration should be done with either a 4 and 7 buffer or a 7 and 10 buffer, depending on sample pH range. Once calibrated, one of the two buffers should be re-checked as if it was a sample. The measured pH should be within ± 0.1 pH unit of actual pH.

Corrective Action Form

QC failures

What QC type failure is involved:

___blank ___known standard ___calibration ___matrix spike ___replicate ___blind ___other

Blank: what is the LOD? ___ What level was detected in the blank? ___

Spikes/replicates: What are the acceptance criteria? ___ Your result? ___

If a matrix spike: Is this a matrix interference? ___ How do you know that? ___

Known standards/blinds: True Value: ___ Acceptance criteria? ___ Your result? ___

Other pertinent information _____

Other problems (equipment malfunctions, etc.)

Symptom(s) (how did you know something was wrong?): _____

Corrective Action Taken

List any activities or checks you performed to identify the source and resolve the problem.

Action/Check Performed	What did you conclude?	Initials	Date
_____	_____	_____	_____
_____	_____	_____	_____
_____	_____	_____	_____
_____	_____	_____	_____
_____	_____	_____	_____

Resolution

Date: _____

Briefly document how you know this problem has been corrected. What changes have you made to prevent it from recurring?

